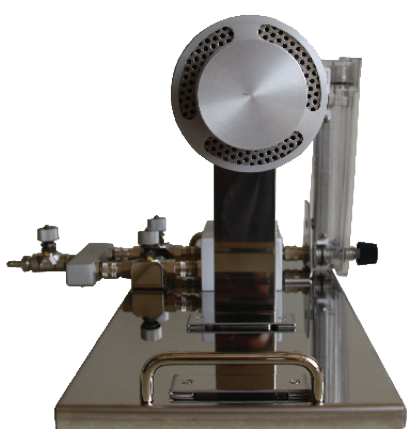
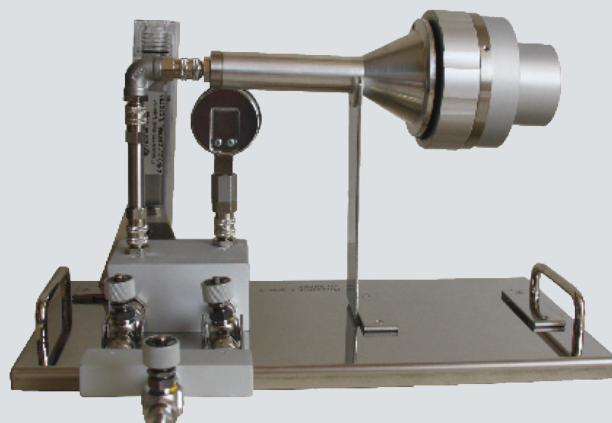
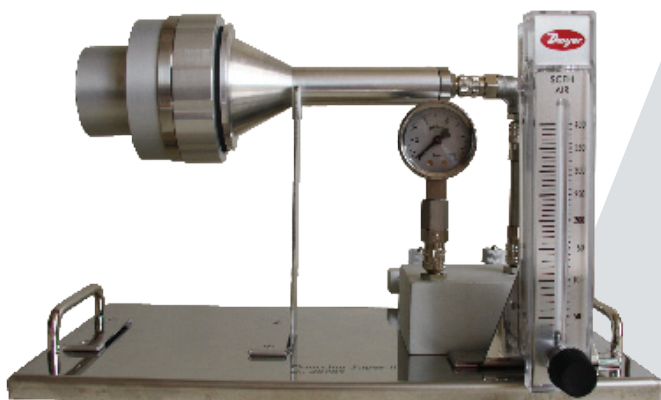


'SAS PINOCCHIO SUPER II' CLEANING AND STERILISATION PROCEDURES



In order to sterilise/disinfect SAS PINOCCHIO SUPER II, you should follow these recommendations:

- 1. Sterilisation with autoclave:** All parts can be autoclaved, with the only exception of the manometer and pressure gauge. Please consider that both pressure gauge and flow meter are not in contact with the sampled air that so their sterilization is not necessary. Dismantle the instrument and sterilise the parts using paper/plastic sterilisation bags for an autoclave. Please ensure that the autoclave you're using has a vacuum pump.
- 2. Sterilisation with irradiation:** All parts can be irradiated. Dismantle the instrument and sterilise the parts using paper/plastic or plastic/plastic sterilisation bags for irradiation. Please ensure that the interior of the tubing is sterilised.
- 3. Sterilisation with flame:** Not recommended because it can burn silicone gaskets and darken the aluminum.
- 4. Disinfection with neutral disinfectants:** All parts can be disinfected this way.
- 5. Disinfection with isopropyl alcohol:** Not recommended because the connections and the handles of the instrument are made by nickel-plated brass that is discoloured by IPA.
- 6. Disinfection with chlorine:** Must be avoided because is too aggressive and can lead to discolouration of the whole instrument.



STERILISATION

Sterilisation must be done if the Pinocchio is used in environments such as cleanrooms where the expected result is 0 CFU per cubic meter. If so we recommend you sterilise all tubes that come into contact with the sampled air (Picture A, red path). We can suggest two kind of sterilisation.

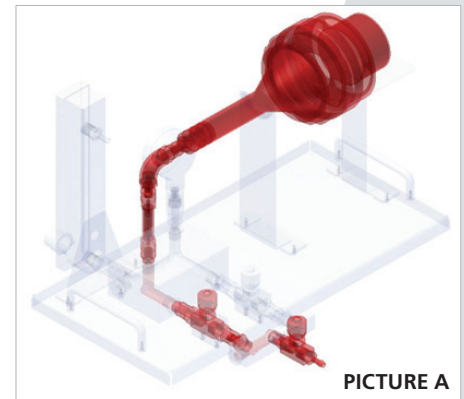
Sterilisation with an autoclave

Is surely the best and simplest. Consider that those parts that can be autoclaved are hollow tubes. As a result, in order to avoid re-contamination of the part it is recommended that you transfer aseptically the sterilised part to the place where the air sampling is taking place. The best way to do that is packing the parts into paper/plastic bags for sterilisation in an autoclave; such as the ones used for sterilising surgical tools.

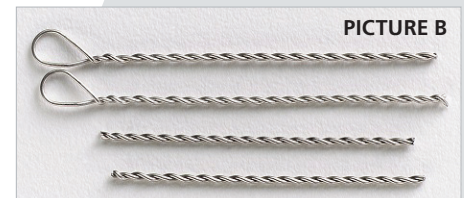
Please consider that is mandatory to use an autoclave with a fractionated vacuum (such as medical class B autoclave) in order to ensure complete sterilisation of hollow tubes inside a bag, and also the drying of the paper after sterilisation to avoid re-contamination. In order to verify periodically the sterilisation of the inside part of the tube it is recommended that you use tiny microbiological indicators, such as needles (Picture B).

Sterilisation with irradiation

Tubes highlighted in Picture A can be also irradiated. It must be ensured that the irradiation process has been validated; especially you must ensure that the internal part of the tubes is reached by the sterilising ray. The same recommendation on the aseptical transfer on sterilisation in autoclave must be followed: Use bags for irradiation.



PICTURE A



PICTURE B

DISINFECTION

As described before, disinfection with IPA and chlorine is not recommended. The first is corrosive for the nickel-plated parts, the second is corrosive for aluminum. So we strongly recommend the use of neutral soaps and disinfectant.

The easiest way to clean the flat parts (such as the base) is simply to use a cloth soaked with the disinfectant or alternatively spray the disinfectant onto the surface and then clean it with a cloth. For all the internal parts we suggest you spray and use small brushes with soft bristles. Be sure to wash the surface properly with distilled water in order to remove the disinfectant. Let the surface dry out.